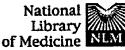
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☐ 1: Biotechnol Appl Biochem. 1996 Feb;23 (Pt 1):67-75.

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Production and characterization of biologically active Ala-Ser-(His)6-Ile-Glu-Gly-Arg-human prolactin (tag-hPRL) secreted in the periplasmic space of Escherichia coli.

Morganti L, Huyer M, Gout PW, Bartolini P.

National Nuclear Energy Commission (IPEN-CNEN)-Cidade Universitaria, Sao Paulo, Brazil.

Human prolactin (hPRL) cDNA was obtained by screening of a pituitary cDNA library with a synthetic 21-mer oligonucleotide and with rat PRL cDNA. For its expression, use was made of a vector, p3SN8, containing tacpromoter-controlled sequences for a bacterial cellulase leader joined to sequences coding for Ala-Ser, a chromatographic affinity site consisting of six histidines and a Factor Xa cleavage site. The hPRL cDNA was inserted at the 3' end of the cleavage-site sequences. Expression in Escherichia coli led to secretion in the periplasmic space of a fully bioactive hPRL variant constituting authentic hPRL with a peptide tag, i.e. Ala-Ser-(His)6-Ile-Glu-Gly-Arg, at its N-terminal. This tag-hPRL could be rapidly and efficiently purified by metal-chelate affinity chromatography. The correct processing and quality of tag-hPRL was monitored by SDS/PAGE, Western-blot analysis, immunoassay and Nb2-lymphoma-cell bioassay. Treatment with Factor Xa for tag removal was only partially successful. Periplasmic secretion of tag-hPRL of the order of 0.7 micrograms/ml per A600 unit and one-step purification indicate feasibility for tag-hPRL production for in vitro diagnostic and research applications. This is the first report describing periplasmic secretion of a bioactive form of hPRL.

PMID: 8867898 [PubMed - indexed for MEDLINE]

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☐ 1: Biotechnol Appl Biochem. 1998 Feb;27 (Pt 1):63-70.

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Synthesis and characterization of recombinant, authentic human prolactin secreted into the periplasmic space of

Escherichia coli.

Morganti L, Soares CR, Affonso R, Gout PW, Bartolini P.

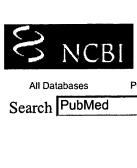
Biotechnology Department, National Nuclear Energy Commission (IPEN-CNEN), Cidade Universitaria, Sao Paulo, Brazil.

Recombinant, fully bioactive, authentic human prolactin (aut-hPRL) has been synthesized in transformed Escherichia coli HB2151 bacteria in a soluble, non-glycosylated form, which is secreted into the bacterial periplasm. Use was made of a bacterial expression vector, containing tac promoter-controlled sequences for the translation enhancer from bacteriophage T7 gene 10, and for a cellulase leader peptide from Cellulomonas fimi joined to sequences coding for aut-hPRL. This vector was derived from a previously described vector containing sequences of an hPRL variant, tag-hPRL (containing a 12-amino-acid peptide tag at the Nterminal end), using site-specific mutagenesis to delete the tag sequence. SDS/PAGE, partial N-terminal amino acid sequence analysis, Western blot analysis and Nb2 lymphoma cell in vitro bioassay indicated correct processing of the hormone. Periplasmic secretion of aut-hPRL, as measured by immunoassay, was relatively low (approx. 0.08 microgram/ml per A600 unit), in contrast to that of tag-hPRL which was approximately 8-fold higher, apparently a consequence of the tag sequence. This is the first report describing periplasmic secretion of biologically active, authentic hPRL.

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Versatile epitope tagging vector for gene expression in

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Hosfield T, Lu Q. Stratagene Cloning Systems, Inc., LaJolla, CA, USA.

mammalian cells.

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☐ 1: Biotechniques. 1998 Aug;25(2):306-9.

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We have constructed an epitope-tagging vector, pCMV-Tag1, for gene expression in mammalian cells. This vector, which allows for N-terminal, Cterminal and internal tagging of the gene product of interest with the FLAG and/or c-myc epitopes, enables researchers to rapidly and efficiently characterize gene products in vivo.

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